

Synthesis of 1,2-*trans*-2-Acetamido-2-deoxyhomoiminosugars

Yves Blériot,^{*,†} Anh Tuan Tran,[‡] Giuseppe Prencipe,[‡] Yeri Jagadeesh,[†] Nicolas Auberger,[†] Sha Zhu,[‡] Charles Gauthier,[†] Yongmin Zhang,[‡] Jérôme Désiré,[†] Isao Adachi,[§] Atsushi Kato,[§] and Matthieu Sollogoub^{*,‡}

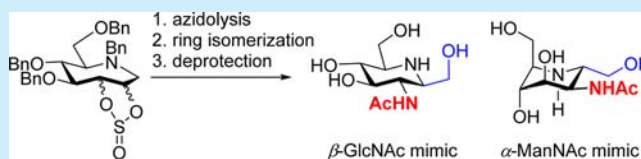
[†]Glycochemistry Group of “Organic Synthesis” Team, Université de Poitiers, UMR-CNRS 7285 IC2MP, Bât. B28, 4 rue Michel Brunet, TSA 51106, 86073 Poitiers Cedex 9, France

[‡]Sorbonne Universités, UPMC Univ Paris 06, Institut Universitaire de France, UMR-CNRS 8232, IPCM, LabEx MiChem, F-75005 Paris, France

[§]Department of Hospital Pharmacy, University of Toyama, 2630 Sugitani, Toyama 930-0194, Japan

Supporting Information

ABSTRACT: The first synthesis of 1,2-*trans*-homoiminosugars devised as mimics of β -D-GlcNAc and α -D-ManNAc is described. Key steps include a regioselective azidolysis of a cyclic sulfite and a β -amino alcohol skeletal rearrangement applied to a polyhydroxylated azepane. The β -D-GlcNAc derivative has been coupled to serine to deliver an iminosugar C-amino acid. The two homoiminosugars demonstrate moderate glycosidase inhibition.



Glycosidase inhibitors are enjoying much interest as they find applications in an increasing number of therapies.¹ Hexosaminidases that trim *N*-acetyl- β -D-glucosamine (GlcNAc) from glycoconjugates are of high therapeutic interest, being involved in several human pathologies including allergy,² osteoarthritis,³ Parkinson's⁴ and Alzheimer's⁵ diseases. Iminosugars, i.e. sugars analogues in which the endocyclic oxygen has been replaced by nitrogen, constitute the most promising class of glycosidase inhibitors. Among the most potent β -hexosaminidase inhibitors, we can mention the naturally occurring pochonicine (1),⁶ siastatin B (2),⁷ nagstatin (3),⁸ the synthetic pyrrolidines LABNAc (4),⁹ ADMDP-acetamide (5),¹⁰ polyhydroxylated proline amide 6,¹¹ azepane (7),¹² and piperidines such as IFGNAc (8),¹³ DNJNAc (9),¹⁴ and DGJNAc (10)¹⁵ (Figure 1). Introduction of structural diversity in compounds 9 and 10, to possibly increase their potency and

selectivity, have mainly focused on the functionalization of the endocyclic nitrogen,¹⁶ the ring hydroxyl groups,¹⁷ or the acetamido group.¹⁸ Introduction of a stereochemically defined and chemically stable pseudoanomeric substituent *cis* to the C-2 substituent of the piperidine ring has been achieved and could constitute a promising alternative.¹⁹ We would like to report herein, and in parallel to our previous paper, a synthetic strategy allowing access to the complementary 1,2-*trans* homo-2-acetamido-1,2-dideoxy iminosugars, illustrated by the synthesis of β -homo-2-acetamido-1,2-dideoxyojirimycin (β -HNJNAc, 11) and α -homo-2-acetamido-1,2-dideoxy-mannoijirimycin (α -HMJNAc, 12) (Figure 1).

The ring isomerization of seven-membered polyhydroxylated azepanes constitutes one strategy among many others to generate new or known piperidine iminosugars.²⁰ This transformation, based on a β -aminoalcohol rearrangement,²¹ requires a free alcohol at the β position and exploits the anchimeric assistance of the nitrogen. Access to 1,2-*trans* 2-acetamido-2-deoxy-homoiminosugars using this approach requires the preliminary *trans* introduction of OH and N₃ functionalities at the respective β and γ positions of the azepane ring. Epoxide azidolysis appears to be a suitable route toward this goal. Epoxidation of known azacycloheptene 13 (see our previous paper) was thus studied (Scheme 1). While *m*-CPBA-, oxone-, or H₂O₂-mediated oxidation gave disappointing results, the procedure developed by Shi²² furnished the 3*R*,4*R* epoxide 14 in an acceptable 54% yield. The stereochemistry of the oxirane ring in 14 was established by comparing the ¹H NMR coupling constants for the protons H-3, H-4, and H-5 that matched those obtained for the

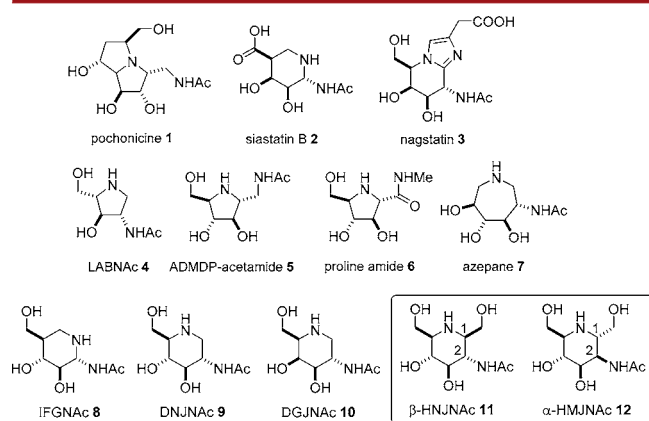
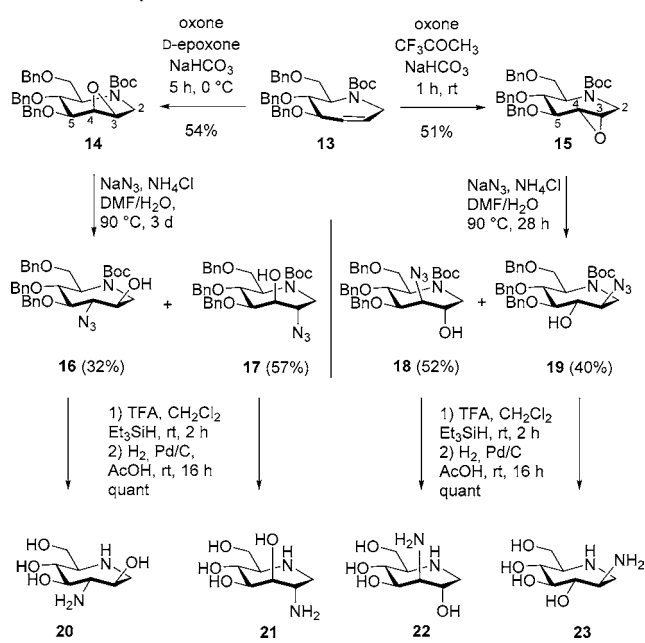


Figure 1. Structure of iminosugars 1–12.

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Scheme 1. Synthesis of Azido Alcohols 16–19



corresponding *N*-Cbz epoxide derivative.²⁴ Additionally, a NOESY experiment supported a *cis* relationship for the H-4 and H-5 protons. In parallel, oxirane-mediated epoxidation using CF₃COCH₃ furnished the diastereomeric 3*S*,4*S* epoxide 15 (51%) along with epoxide 14 (29%). Azidolysis of 14 using NaN₃ and NH₄Cl in DMF/H₂O at 90 °C afforded the required *trans* azido alcohol 16 as the minor product (32%) along with its regioisomer 17 (57%). Similar treatment of epoxide 15 furnished the azido alcohol 18 (52%) along with its regioisomer 19 (40%). Because of the presence of rotamers, the stereochemistry of azido alcohols 16–19 was difficult to elucidate by NMR. Their firm structure assignment was achieved via their deprotection with TFA followed by hydrogenolysis (H₂, Pd/C, AcOH) to give the known tetrahydroxylated aminoazepanes 20–23, the ¹H NMR data for which were in good agreement with the literature.^{23b} The observed regioselectivities for the azidolysis step are similar to those reported in the case of a *N*-Cbz protecting group.²³ While the well-established Fürst–Plattner principle²⁴ of *trans* diaxial epoxide ring-opening usually furnishes major diastereoisomers in the case of piperidines,²⁵ the azepane ring flexibility can be invoked here to explain the lack of regiocontrol during azidolysis. On the basis of the disappointing results above (17% yield for azido alcohol 16 and 26% yield for azido alcohol 18 from compound 13), we concluded that a second round of optimization would be necessary to achieve synthetically useful yields. We anticipated that protection of the corresponding 3,4-*cis* diols as their cyclic sulfites²⁶ could provide the synthetic equivalent of an epoxide, possibly allowing the regioselective introduction of an azide at the γ position. To this end, several asymmetric dihydroxylation conditions were applied to 13 in order to improve the modest level of diastereoselectivity obtained in the case of OsO₄-mediated dihydroxylation (Table 1).

The best conversion and diastereoselectivities were obtained with the modified Sharpless asymmetric dihydroxylation (SAD) method²⁷ using AD-mix α and β , respectively, with 0.2 equiv of ligand (entries 4 and 7) that afforded the bottom-face diol 24 and the top-face diol 25 in 65% and 58% yields, respectively. As

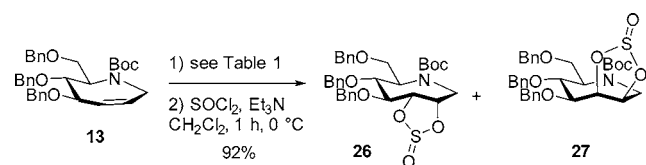
Table 1. Dihydroxylation of Azacycloheptene 13

entry	conditions	24:25 ratio ^a
1	OsO ₄ , acetone/H ₂ O	2:1
2	α -AD-mix, <i>t</i> -BuOH/H ₂ O	1.5:1
3	modified SAD, 0.05 equiv of ligand I, ^b <i>t</i> -BuOH/H ₂ O	2.5:1
4	modified SAD, 0.2 equiv of ligand I, ^b <i>t</i> -BuOH/H ₂ O	2.9:1
5	β -AD-mix, <i>t</i> -BuOH/H ₂ O	1:1.4
6	modified SAD, 0.05 equiv of ligand II, ^b <i>t</i> -BuOH/H ₂ O	1:1.5
7	modified SAD, 0.2 equiv of ligand II, ^b <i>t</i> -BuOH/H ₂ O	1:1.9

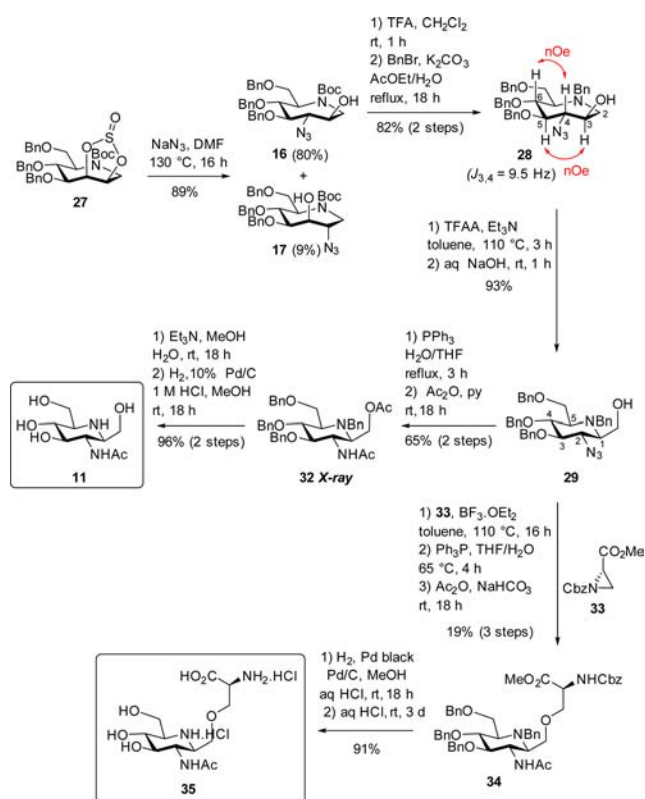
^aThe ratio of diols 24/25 was determined by ¹H NMR by integrating the protons of the Boc group. ^bLigand I = [(DHQD)₂PHAL]; ligand II = [(DHQD)₂PHAL].

these diols were found to be difficult to separate on a large scale, the mixture of diols obtained after dihydroxylation was directly treated with thionyl chloride and Et₃N to furnish the separable sulfites 26 and 27 in an excellent 92% yield (Scheme 2). Because of the presence of a stereogenic sulfur atom, 26 and 27 were both obtained as a mixture of diastereoisomers that were not separated and directly used in the next step.

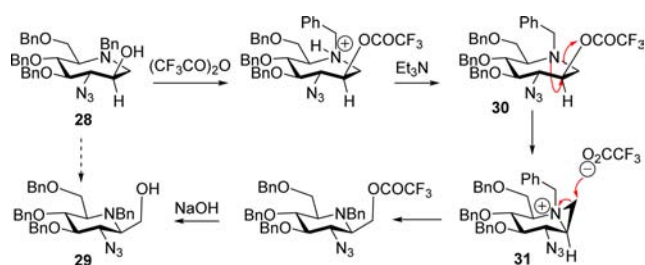
Scheme 2. Synthesis of Cyclic Sulfites 26 and 27



With the synthesis of β -HNJNac (11) in mind, we took advantage of the cyclic sulfite 27 to study its azidolysis (Scheme 3).²⁸ Treatment of 27 with LiN₃ (20% solution in water) at 130 °C provided the desired azido alcohol 16 (57%). A significant amount of diol 25 (25%) was also recovered probably arising from cyclic sulfite hydrolysis. Switching to neat NaN₃ at 130 °C furnished the required azido alcohol 16 in 80% yield (42% from 13) along with its regioisomer 17 (9%). Removal of the Boc group in 16 with TFA followed by *N*-benzylation furnished the *N*-benzyl azepane 28 (82% over two steps), which was characterized by a large coupling constant between the *trans* H-3 and H-4 protons ($J = 9.5$ Hz) and NOE contacts between H-3 and H-5 and between H-4 and H-6. The β -amino alcohol rearrangement of 28 under Mitsunobu conditions proved unsatisfactory, affording the piperidine 29 in low yield. Use of TFAA²⁹ was beneficial as it furnished piperidine 29 in excellent yield (93%) after ester hydrolysis. For this transformation, we propose a mechanism (Scheme 4) in which the free alcohol in 28 is activated as its trifluoroacetate ester 30 and displaced by the endocyclic nitrogen, generating a fused piperidine–aziridinium ion 31. The released trifluoroacetate ion then attacks the methylene carbon of the aziridinium ion and displaces the ammonium group to furnish the piperidine 29 after ester hydrolysis. ¹H NMR analysis of 29 ($J_{1,2} = 10.0$ Hz, $J_{3,4} = J_{4,5} = 8.0$ Hz) confirmed its β -D-*gluco* configuration. Conversion of the azide functionality into an acetamide under standard conditions (PPh₃, THF/H₂O then Ac₂O, py) gave the crystalline diacetylated derivative 32 (61%), the crystal

Scheme 3. Synthesis of β -HNJNAC 11 and Conjugate 35

Scheme 4. Proposed Mechanism for the Formation of Piperidine 29 from 28



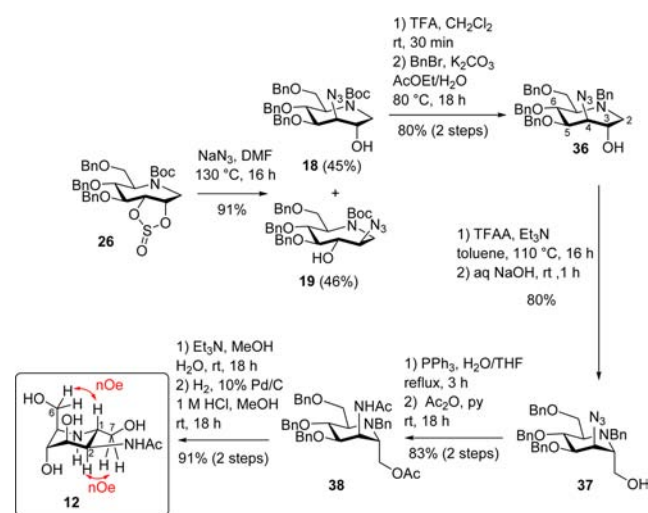
structure of which was solved (Figure 2). O-Deacetylation with Et_3N in MeOH followed by hydrogenolysis under mild acidic



Figure 2. X-ray crystallography of compound 32 (CCDC 1015486).

conditions furnished the target β -HNJNAC 11 (Scheme 3).³⁰ To demonstrate the potential of these derivatives as “iminosugar C-glycosyl donors”, homoiminosugar 29 was coupled to the serine precursor 33³¹ to yield the corresponding iminosugar amino acid precursor 34 after functional group interconversion from azide to acetamide. Hydrogenolysis followed by ester hydrolysis provided the homoiminosugar amino acid 35 (Scheme 3).

To further exemplify our methodology, we synthesized a mannose analogue, namely the α -homo-2-acetamido-1,2-dideoxymannojirimycin (α -HMJNAC, 12). Azidolysis of the cyclic sulfite 26 under the conditions depicted above furnished the azido alcohol 18 in 45% yield (27% from 13) along with its regioisomer 19 (46%). Replacement of the Boc group by the electron-donating benzyl group yielded azepane 36, as characterized by a small coupling constant between the *cis* H-4 and H-5 protons ($J = 1.5$ Hz) and a correlation in the COSY spectrum between H-3 and the free OH. Treatment of 36 with TFAA in toluene at 110 °C yielded the piperidine 37 in 80% yield after ester hydrolysis. Introduction of the acetamide group in 37 uneventfully provided piperidine 38 in 83% yield. O-Deacetylation followed by hydrogenolysis under mild acidic conditions yielded the target α -HMJNAC 12. The *trans* relationship between the NHAc function and the pseudoanomeric CH_2OH group is confirmed by a NOESY cross-correlation between H-2 and H-7 (Scheme 5). NMR analysis

Scheme 5. Synthesis of α -HMJNAC 12

indicated that, in CD_3OD , β -HNJNAC (11) adopts a β -glucose-like ${}^4\text{C}_1$ -type conformation ($J_{2,3} = 10.1$ Hz, $J_{3,4} = 8.8$ Hz, $J_{4,5} = 9.7$ Hz) and α -HMJNAC (12) an inverted ${}^1\text{C}_4$ chair ($J_{2,3} = 3.5$ Hz, $J_{3,4} = 3.5$ Hz, $J_{4,5} = 3.5$ Hz and H-6/H-1 NOESY cross-correlation).

The β -HNJNAC (11) and α -HMJNAC (12) were assayed on a panel of β -N-acetylhexosaminidases from human placenta, bovine kidney, HL-60, Jack bean, and *Aspergillus oryzae* and α -N-acetylgalactosaminidase from chicken liver (Table 2). Surprisingly, β -HNJNAC (11) is only a moderate inhibitor of

Table 2. Concentration (in μM) of Iminosugars 11 and 12 Giving 50% Inhibition of Various Glycosidases (IC_{50})

enzyme	11	12
β -N-acetylhexosaminidase		
human placenta	72	302
bovine kidney	65	624
HL-60	88	394
Jack bean	41	95
<i>Aspergillus oryzae</i>	NI ^a	NI
α -N-acetylgalactosaminidase chicken liver	NI	NI

^aNI: no inhibition (less than 50% inhibition at 1000 μM).

β -N-acetylhexosaminidases. As expected, α -HMJNac (**12**), which displays a different configuration for two hydroxyl groups compared to the parent *gluco*-configured substrate, is a poor inhibitor of these enzymes.

In summary, the first synthesis of 1,2-*trans* homoiminosugars derived from GlcNAc and ManNAc bearing a pseudoanomeric CH₂OH group is reported exploiting a β -amino alcohol rearrangement applied to a seven-membered iminosugar. Use of a cyclic sulfite derivative as an epoxide equivalent and its azidolysis proved beneficial to access the azepane precursor necessary for the ring-contraction step. This work has produced novel structures that could be used as probes in the field of β -N-acetylhexosaminidases.

■ ASSOCIATED CONTENT

Supporting Information

Experimental details, NMR spectra, and X-ray crystallography data. This material is available free of charge via the Internet at <http://pubs.acs.org>.

■ AUTHOR INFORMATION

Corresponding Authors

*E-mail: yves.blériot@univ-poitiers.fr.

*E-mail: matthieu.sollogoub@upmc.fr.

Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) (a) Horne, G.; Wilson, F. X. *Prog. Med. Chem.* **2011**, *50*, 135–176. (b) Horne, G.; Wilson, F. X.; Tinsley, J. D.; Williams, H.; Storer, R. *Drug Discovery Today* **2011**, *16*, 107–118.
- (2) Reese, T. A.; Liang, H.-E.; Tager, A. M.; Luster, A. D.; Van Rooijen, N.; Voehringer, D.; Locksley, R. M. *Nature* **2007**, *447*, 92–96.
- (3) Liu, J.; Shikhman, A. R.; Lotz, M. K.; Wong, C. H. *Chem. Biol.* **2001**, *8*, 701–711.
- (4) Groves, J. A.; Lee, A.; Yildirim, G.; Zachara, N. E. *Cell Stress Chaperones* **2013**, *18*, 535–558.
- (5) Cecioni, S.; Vocadlo, D. J. *Curr. Opin. Chem. Biol.* **2013**, *17*, 719–728.
- (6) Usuki, H.; Toyo-oka, M.; Kanzaki, H.; Okuda, T.; Nitoda, T. *Bioorg. Med. Chem.* **2009**, *17*, 7248–7253.
- (7) Umezawa, H.; Aoyagi, T.; Komiyama, T.; Morishima, H.; Hamada, M.; Takeuchi, T. *J. Antibiot.* **1974**, *27*, 963–969.
- (8) Aoyama, T.; Naganawa, H.; Suda, H.; Uotani, K.; Aoyagi, T.; Takeuchi, T. *J. Antibiot.* **1992**, *45*, 1557–1558.
- (9) Rountree, J. S. S.; Butters, T. D.; Dwek, R. A.; Asano, N.; Ikeda, K.; Evinson, E. L.; Nash, R. J.; Fleet, G. W. J. *Tetrahedron Lett.* **2007**, *48*, 4287–4291.
- (10) Liang, P.-H.; Cheng, W.-C.; Lee, Y.-L.; Yu, H.-P.; Wu, Y.-T.; Lin, Y.-L.; Wong, C.-H. *ChemBioChem* **2006**, *7*, 165–173.
- (11) Ayers, B. J.; Glawar, A. F. G.; Martínez, R. F.; Ngo, N.; Liu, Z.; Fleet, G. W. J.; Butters, T. D.; Nash, R. J.; Yu, C.-Y.; Wormald, M. R.; Nakagawa, S.; Adachi, I.; Kato, A.; Jenkinson, S. F. *J. Org. Chem.* **2014**, *79*, 3398–3409.
- (12) (a) Li, H.; Marcelo, F.; Bello, C.; Vogel, P.; Butters, T. D.; Sollogoub, M.; Rauter, A. P.; Blériot, Y. *Bioorg. Med. Chem.* **2009**, *17*, 5598–5604. (b) Marcelo, F.; He, Y.; Yuzwa, S. A.; Nieto, L.; Jiménez-Barbero, J.; Sollogoub, M.; Vocadlo, D. J.; Davies, G. J.; Blériot, Y. J.

Am. Chem. Soc. **2009**, *131*, 5390–5392. (c) Mondon, M.; Hur, S.; Vadlamani, G.; Rodrigues, P.; Madden, Z.; Oliver, A.; Mark, B. L.; Vocadlo, D. J.; Blériot, Y. *Chem. Commun.* **2013**, *49*, 10983–10985.

(13) Shitara, E.; Nishimura, Y.; Kojima, F.; Takeuchi, T. *Bioorg. Med. Chem.* **1999**, *7*, 1241–1246.

(14) (a) Fleet, G. W. J.; Fellows, L. E.; Smith, P. W. *Tetrahedron* **1987**, *43*, 979–990. (b) Fleet, G. W. J.; Smith, P. W.; Nash, R. J.; Fellows, L. E.; Parekh, R. B.; Rademacher, T. W. *Chem. Lett.* **1986**, 1051–1054. (c) Glawar, A. F. G.; Best, D.; Ayers, B. J.; Miyauchi, S.; Nakagawa, S.; Aguilar-Moncayo, M.; Garcia, F. J. M.; Ortiz, M. C.; Crabtree, E. V.; Butters, T. D.; Wilson, F. X.; Kato, A.; Fleet, G. W. J. *Chem.—Eur. J.* **2012**, *18*, 9341–9359.

(15) Best, D.; Chairatana, P.; Glawar, A. F. G.; Crabtree, E.; Butters, T. D.; Wilson, F. X.; Yu, C.-Y.; Wang, W.-B.; Jia, Y.-M.; Adachi, I.; Kato, A.; Fleet, G. W. J. *Tetrahedron Lett.* **2010**, *51*, 2222–2224.

(16) (a) Ho, C.-W.; Papat, S. D.; Lei, T.-W.; Tsai, K.-C.; Ho, M.-J.; Chen, W.-H.; Yang, A.-S.; Lin, C.-H. *Chem. Biol.* **2010**, *5*, 489–497. (b) Steiner, A. J.; Schitter, G.; Stütz, A. E.; Wrodnigg, T. M.; Tarling, C. A.; Withers, S. G.; Mahuran, D. J.; Tropak, M. B. *Tetrahedron: Asymmetry* **2009**, *20*, 832–835.

(17) Yamaguchi, T.; Blazquez, B.; Heseck, D.; Lee, M.; Llarrull, L. I.; Bogges, B.; Oliver, A. G.; Fisher, J. F.; Mobashery, S. *ACS Med. Chem. Lett.* **2012**, *3*, 238–242.

(18) Stubbs, K. A.; Bacik, J.-P.; Perley-Robertson, G. E.; Whitworth, G. E.; Gloster, T. M.; Vocadlo, D. J.; Mark, B. L. *ChemBioChem* **2013**, *14*, 1973–1981.

(19) During the revision of this manuscript, the group of Wong reported the synthesis of iminosugar C-glycoside analogues of α -D-GlcNAc-1-phosphate; see Hsu, C.-H.; Schelwies, M.; Enck, S.; Huang, L.-Y.; Huang, S.-H.; Chang, Y.-F.; Cheng, T.-J. R.; Cheng, W.-C.; Wong, C.-H. *J. Org. Chem.* **2014**, *79*, 8629–8637.

(20) (a) Poitout, L.; Le Merrer, Y.; Depezay, J.-C. *Tetrahedron Lett.* **1996**, *37*, 1613–1616. (b) Bagal, S. K.; Davies, S. G.; Lee, J. A.; Roberts, P. M.; Russell, A. J.; Scott, P. M.; Thompson, J. E. *Org. Lett.* **2010**, *12*, 136–139.

(21) Métro, X.; Duthion, B.; Gomez Pardo, D.; Cossy, J. *Chem. Soc. Rev.* **2010**, *39*, 89–102.

(22) Lorenz, J. C.; Frohn, M.; Zhou, X.; Zhang, J.-R.; Tang, Y.; Burke, C.; Shi, Y. *J. Org. Chem.* **2005**, *70*, 2904–2911.

(23) (a) Li, H.; Schütz, C.; Favre, S.; Zhang, Y.; Vogel, P.; Sinay, P.; Blériot, Y. *Org. Biomol. Chem.* **2006**, *4*, 1653–1662. (b) Li, H.; Blériot, Y.; Mallet, J.-M.; Rodriguez-Garcia, E.; Vogel, P.; Zhang, Y.; Sinay, P. *Tetrahedron: Asymmetry* **2005**, *16*, 313–319.

(24) Fürst, A.; Plattner, P. A. *Helv. Chim. Acta* **1949**, *32*, 275–283.

(25) Coombs, T. C.; Lishington, G. H.; Douglas, J.; Aubé, J. *Angew. Chem., Int. Ed.* **2011**, *50*, 2734–2737.

(26) (a) Megia-Fernandez, A.; Morales-Sanfrutos, J.; Hernandez-Mateo, F.; Santoyo-Gonzalez, F. *Curr. Org. Chem.* **2011**, *15*, 401–432. (b) Byun, H.-S.; He, L.; Bittman, R. *Tetrahedron* **2000**, *56*, 7051–7091.

(27) Walsh, P. J.; Sharpless, K. B. *Synlett* **1993**, 605–606.

(28) (a) Cakmak, P.; Mayer, M.; Trauner, D. *Nat. Chem.* **2011**, *3*, 543–545. (b) Rejman, D.; Pohl, R.; Dracinsky, M. *Eur. J. Org. Chem.* **2011**, 2172–2187.

(29) Metro, T.-X.; Appenzeller, J.; Gomez Pardo, D.; Cossy, J. *Org. Lett.* **2006**, 3509–3512.

(30) A precursor of this compound has been reported; see: Granier, T.; Vasella, A. *Helv. Chim. Acta* **1998**, *81*, 865–880.

(31) Ryan, D. A.; Gin, D. Y. *J. Am. Chem. Soc.* **2008**, *130*, 15228–15230.